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Museum epigenomics as a Toolbox in Evolutionary Research

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PROJECT DESCRIPTION

Natural history collections harbor a vast reservoir of information on the molecular mechanisms underlying biological diversity, but unlocking these 'hidden treasures' remains a daunting task. Only recently researchers have found ways to access this genetic repository thanks to ground-breaking advancements in next-generation sequencing technology. While museum studies on the spatial and/or temporal variability of nucleotide sequences have greatly advanced our understanding on the molecular basis of phenotypic variation, they have remained largely ignorant on exploring other important drivers of phenotypic diversity such as epigenetic modifications of DNA. The awareness of such lacuna has stimulated the emergence of a new research field, 'museum epigenomics'. Studying the epigenomes of museum samples is however far from straightforward as DNA will typically endure a variety of post-mortem alterations like deamination, fragmentation, loss and exogenous contamination, which all complicate the implementation of standard epigenetic lab protocols. In this project we will study these difficulties in an attempt to make this epigenetic repository more accessible for future research. While there are many different epigenetic modifications that DNA can endure, we will target here specifically DNA methylation. The addition of methyl groups to the DNA back-bone, is a frequently used epigenetic marker and more importantly remains stable over extensive time periods making it specifically useful when studying the epigenome of museum samples.

In this project we specifically aim to:

- outline the 'best-practices' in epigenetic profile assessments of museum collections, and
- ii) illustrate how the exploration of epigenomic landscapes of museum collections can provide valuable ecological and evolutionary insights using two pilot studies.

We will here focus on two different methylation profiling methods, reduced representation bisulphite sequencing (RRBS) and whole genome bisulphite sequencing (WGBS). Both methods have their own pros and cons and the choice of which method to use largely depends on the research question and sampling design. While WGBS covers the entire genome, it is currently still too expensive to screen large numbers of specimens. RRBS, on the other hand, is highly cost efficient and allows to process hundreds of specimens within a single project, yet bears the risk of missing out important regions as it only covers a fraction of the genome. We will explore various lab protocols to define the 'best practices' for constructing next-generation sequencing libraries of historical museum samples and assess to what extent various sample characteristics (i.e. age, storage condition, DNA fragmentation, DNA quantity) determine the successful implementation of methylomics in natural history collections. To exemplify how natural history museum collections can aid in exploring the role of epigenetics in ecology and evolution, we will make use of two museum collections of the RBINS. The first one covers a unique adaptive radiation of Calosoma beetles at the Galápagos islands, where representatives of this genus radiated repeatedly into a highland ('long-winged') and lowland ('short-winged') ecotype along an altitudinal gradient replicated on all major islands. This radiation is characterized by a clear divergence gradient between highland and lowland species that neatly correlates with island age. This unique museum collection allows us to explore which role methylation plays in adaptive evolution, especially at the early stages of divergence. The second museum collection highlights the shell polymorphism of the intertidal periwinkle Tectarius striatus sampled at the Azores. Two shell morphotypes within this species co-occur at a microgeographical scale but display a non-random distribution across the landscape as their shell morphology strongly hinges on the extent of wave exposure they endure. By linking genetic, epigenetic and morphological variation we will explore the role of genetic determinism versus phenotypic plasticity in shell variation within this species.



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This project strives to have a direct impact on collection management by allowing researchers access to a new 'data layer' obtained from collection material, an epigenomic archive, complementary to existing morphological and genomic archives, and stimulate new collection-based research lines in various scientific fields.

CONTACT INFORMATION

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